

# A Novel Approach to Internal Standardization in LC/MS/MS Analysis

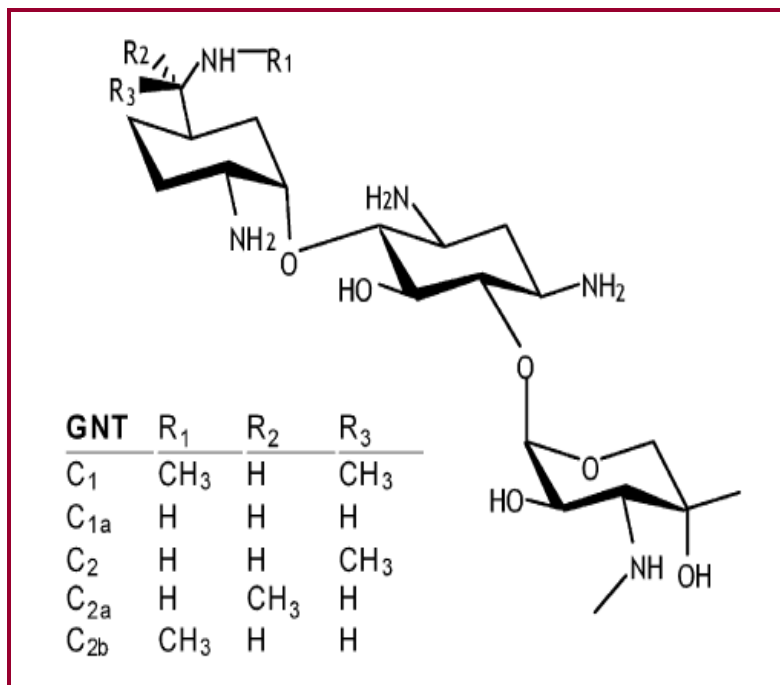
## SENSITIVE LC/MS/MS ANALYSIS OF GENTAMICIN

**CACO-PBSS Mini-Symposium:**  
Bioanalytical and Analytical Applications  
and Problem Investigation Case Studies  
August 10, 2012

# Background & Introduction

**Gentamicin** is an aminoglycoside antimicrobial agent produced by fermentation of *Micromonospora purpurea* or *M. echinospora*. It has a wide spectrum of antimicrobial activity. Gentamicin is not a single molecule but a complex of three major and several minor components. Gentamicins  $C_1$ ,  $C_{1a}$  and  $C_2$  are the three major components of the drug complex. The  $C_2$  component consists of two stereoisomers ( $C_2$  and  $C_{2a}$ ).

## Structures of Gentamicin Components



## Molecular Weights and Approximate Composition

Gentamicin C <sub>1a</sub>	MW 449.5	10-35%
Gentamicin C <sub>2</sub> & C <sub>2a</sub>	MW 463.6	25-55%
Gentamicin C <sub>1</sub>	MW 477.6	25-50%

# The Analytical Challenge

- Gentamicin is associated with severe side effects
- Use is subject to therapeutic drug monitoring, especially in patients with renal impairment
- Product is being developed for subcutaneous injection at surgical incision sites to prevent post surgery infections
  - High systemic exposure not intended and hopefully avoided
  - Circulating plasma levels likely to be very low relative to IM, IV or oral routes of dosing
- LLOQ of 1.00 ng/mL requested
- Very low sample volumes are available
- Gentamicin is co-administered with Vancomycin requiring sample volume for a separate assay

# Approaches in the Literature

## Pharmaceutical Product Analyses:

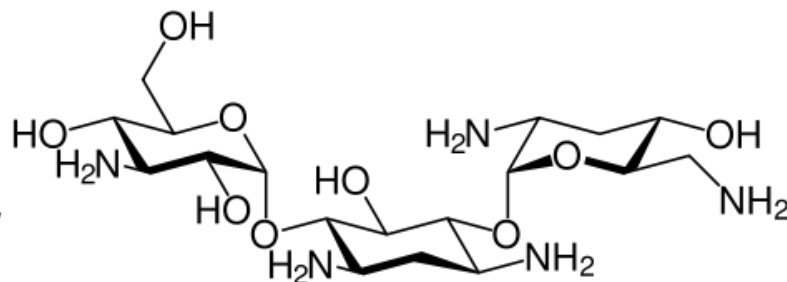
- USP monograph. Pre-column labeling with o-phthalaldehyde (OPA). Methodology makes a somewhat flawed assumption for equal response of components as they have different number of reactive primary amines.
- Similar OPA derivative with micellar electrokinetic chromatography and UV @ 340 nm
- HPLC 2012 poster by Thermo with ion-paired RP-HPLC (HFBA, TFA) charged aerosol detection
- Reversed phase ion-pairing (octane sulfonic acid) pulsed electrochemical detection
- Reversed phase ion-pairing TFA and methanol and electrospray LC/MS/MS

## Bioanalytical Analyses of Plasma or Tissues:

- SPE with on-column derivatization with 1-fluoro-2,4-dinitrobenzene RP-HPLC ultraviolet absorbance at 365 nm
- Derivatization with phenyl isocyanate and RP-HPLC UV
- Fluorenylmethyloxycarbonyl chloride (FMOC) derivatization, UV 265 nm or fluorescence
- Tissues: HPLC with post column derivatization with OPA and fluorescence detection limits 400 ng/gm
- Hydrophilic interaction chromatography (HILIC) with MS/MS detection

# Our Approach

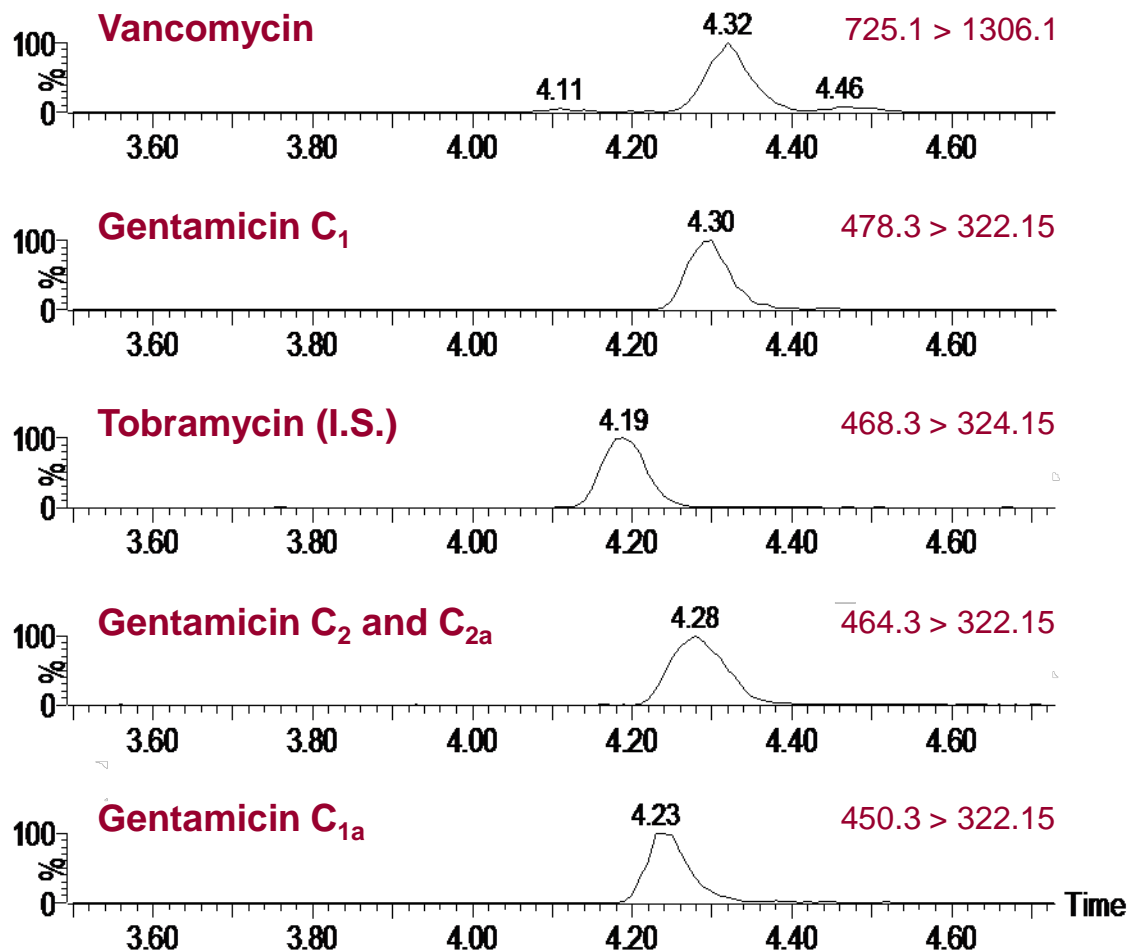
- Common themes: Ion Pairing or Derivatization
- No precedent in literature to achieve required 1.00 ng/mL LLOQ using small plasma volumes
- Initial approach = LC/MS/MS
- No isotopically labeled I.S. would ever be available
  - Even if a single isoform could be synthesized would it track the other isoforms?
  - Spent considerable time and effort with an analogue I.S. Tobramycin (MW 467.51)
- Ion – Paired SPE with HFBA pretreatment on micro-elution C18 followed by LC/MS/MS
- Decided to quantify C<sub>2</sub> and C<sub>2a</sub> together (same MW and not readily resolved chromatographically)



**Tobramycin**

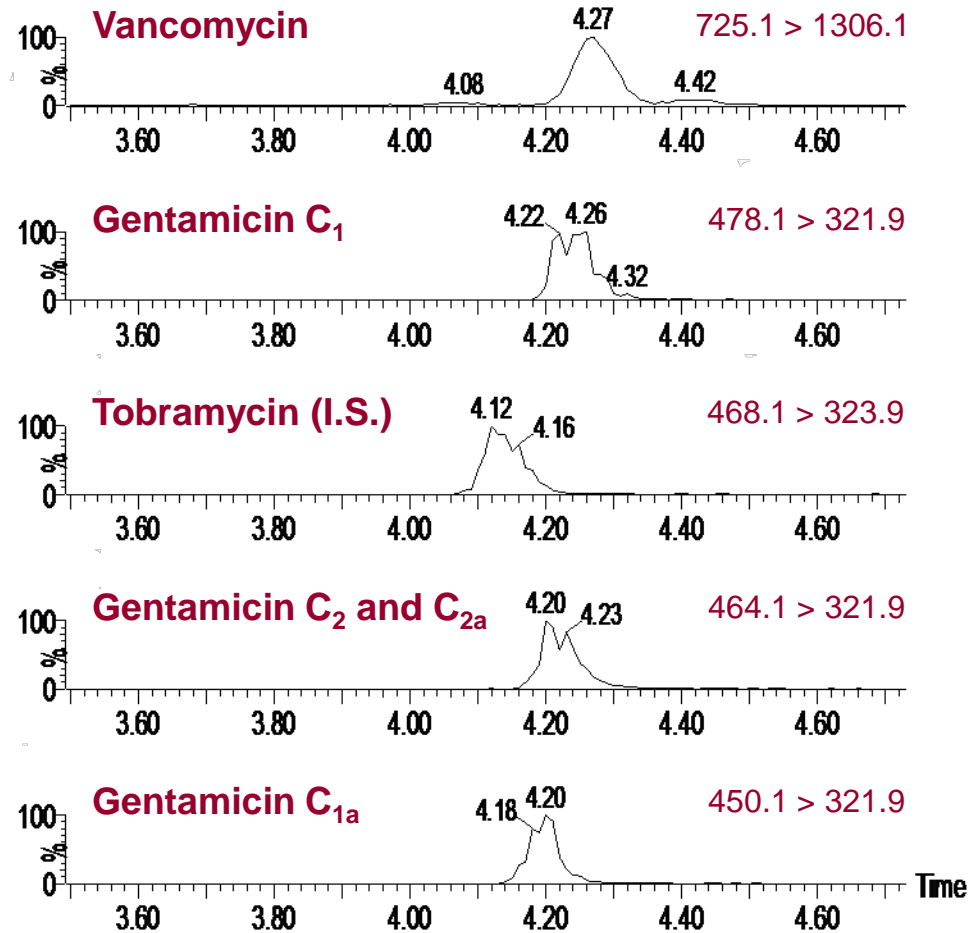
# Initial Results

- Made considerable progress
- Sensitivity looked promising
- HFBA modifier in a HILIC-type separation
- Attempted to include Vancomycin



# Results Over Time

- Vancomycin was successfully validated using an analogue I.S. (Ristomycin). LLOQ was 0.500 ng/mL using 20  $\mu$ L of plasma.
- Gentamicin exhibited poor ionization stability in the source
- Good progress, but intuition suggested the technique would not validate per FDA Guidance
- Other options were pursued



# How About Derivatization with LC/MS/MS?

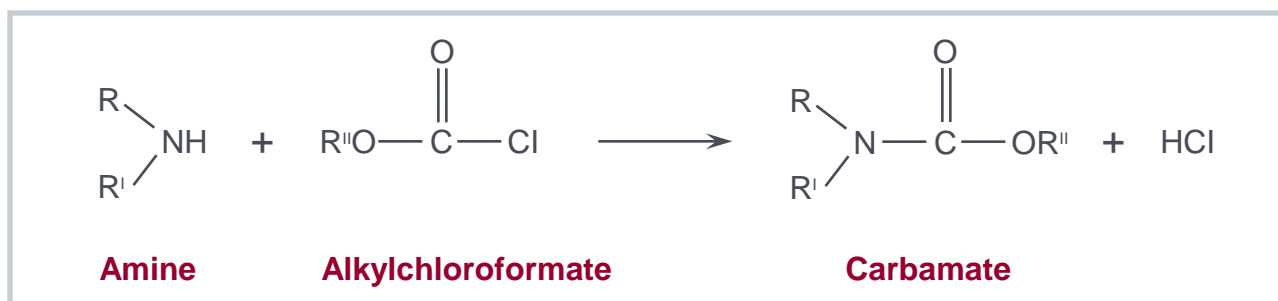
**For this application, what are the attributes of a good derivative?**

1. Needs to tag all 5 amines efficiently and robustly
2. Need a stable derivative, one not affected by moisture during formation or stability post reaction
3. Pre-column derivatization for simplicity... let's not add unnecessary mechanical complexity
4. Derivative should impart added hydrophobicity allowing more selective clean-up and chromatographic options
5. Bigger, but not too big! With 5 amines to tag, things could get out of hand quickly.
6. Improves LC/MS/MS sensitivity and ionization consistency – potentially new product ions and greater yield could vastly improve overall response
7. I.S. tracking – could the correct derivative make an analogue I.S. track better?
8. Manageable excess – a derivatization reagent for which excess reagent would either not be a problem or be readily eliminated



# We Gave it a Try...

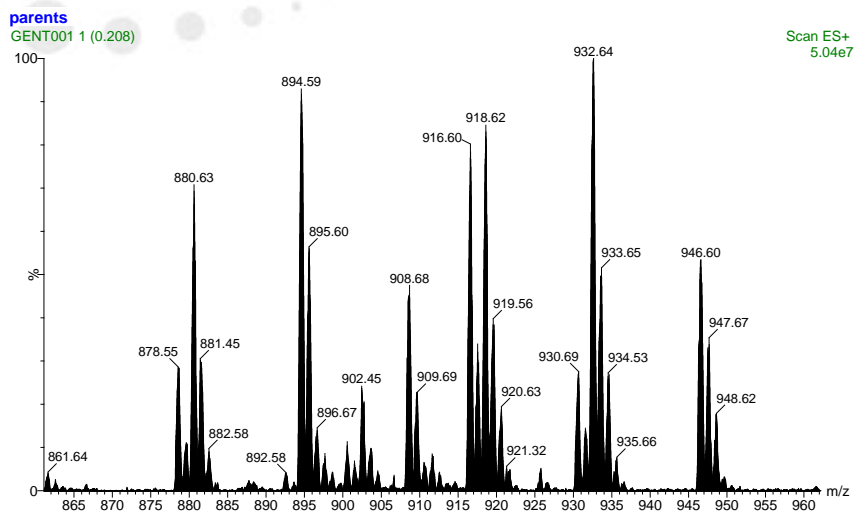
- Given the attributes of a good derivative (previous slide), could all the criteria be met using alkyl chloroformates?
- Reaction on primary and secondary amines:



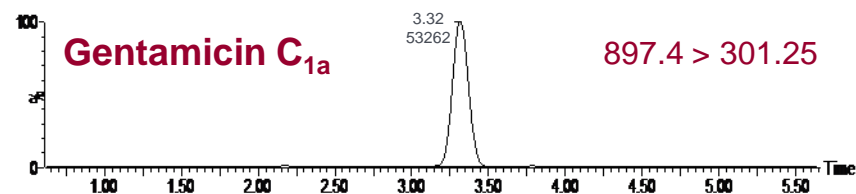
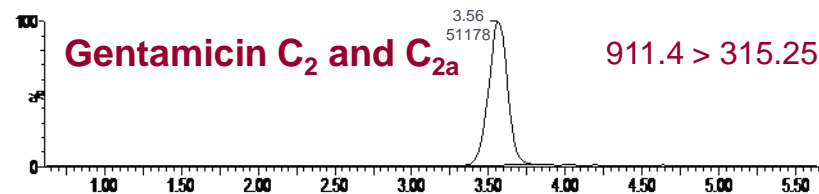
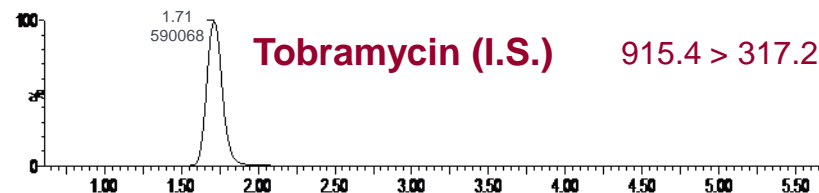
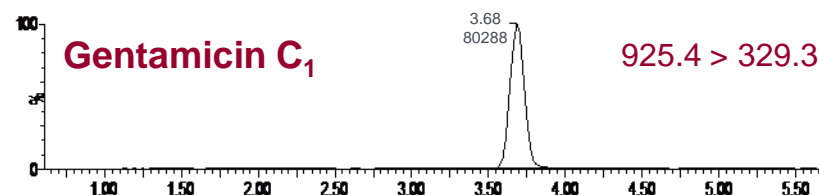
- Explored both ethyl chloroformate and propyl chloroformate
  - Reaction is base-catalyzed and works in aqueous phase
  - Acetone is a good co-solvent
- Have previous experience with the derivatization of Amikacin (related amino glycoside) using ethyl chloroformate
- Only obvious initial disadvantage was ionization - chloroformate derivatives of Gentamicin can find sodium where it doesn't exist!

# Initial Outcome

## Direct MS Infusion of PCF Reaction Product



PCF derivatives of isomeric forms  $C_2$  and  $C_{2a}$  were not chromatographically resolved and were treated as a single compound:



What masses can we expect for a 5 X PCF reaction?

Gentamicin Form:	$C_{1a}$	$C_2$ & $C_{2a}$	$C_1$
Original Parent	450	464	478
Derivative (no adduct)	880	894	908
Sodium Adduct	902	916	930
Potassium Adduct	936	933	947
<b>Ammonium Adduct</b>	<b>897</b>	<b>911</b>	<b>925</b>

Extracted standard representing 50 ng/mL total Gentamicin

# Proof of Concept... Does it Work?

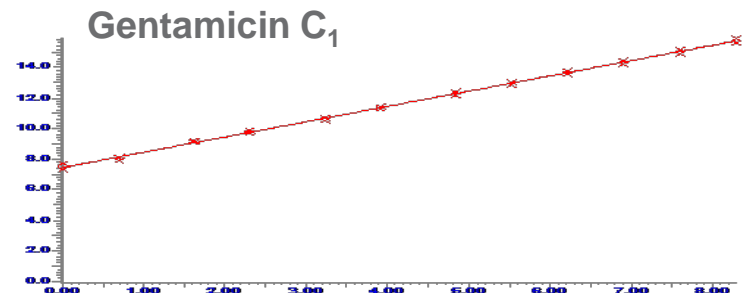
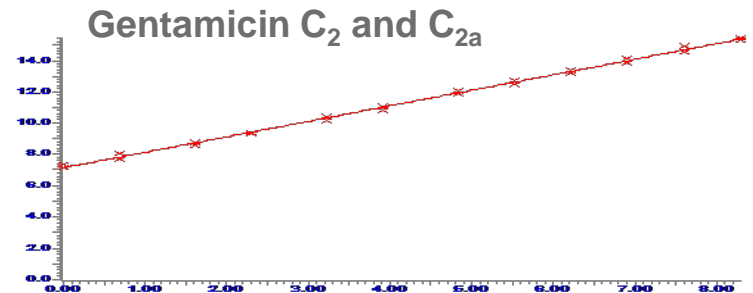
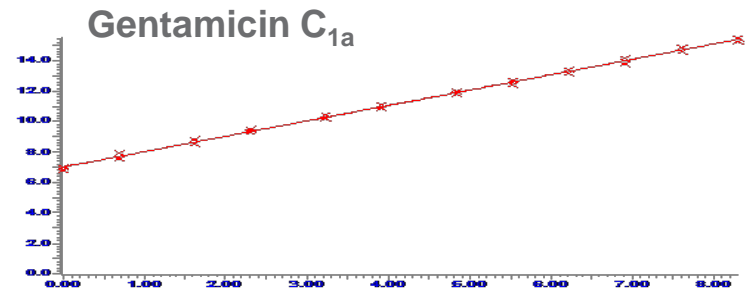
- Tested extraction of 50  $\mu\text{L}$  of plasma and calibration range from 1.00 to 4,000 ng/mL
- Calculations by external standard showed promise!

## GOOD NEWS

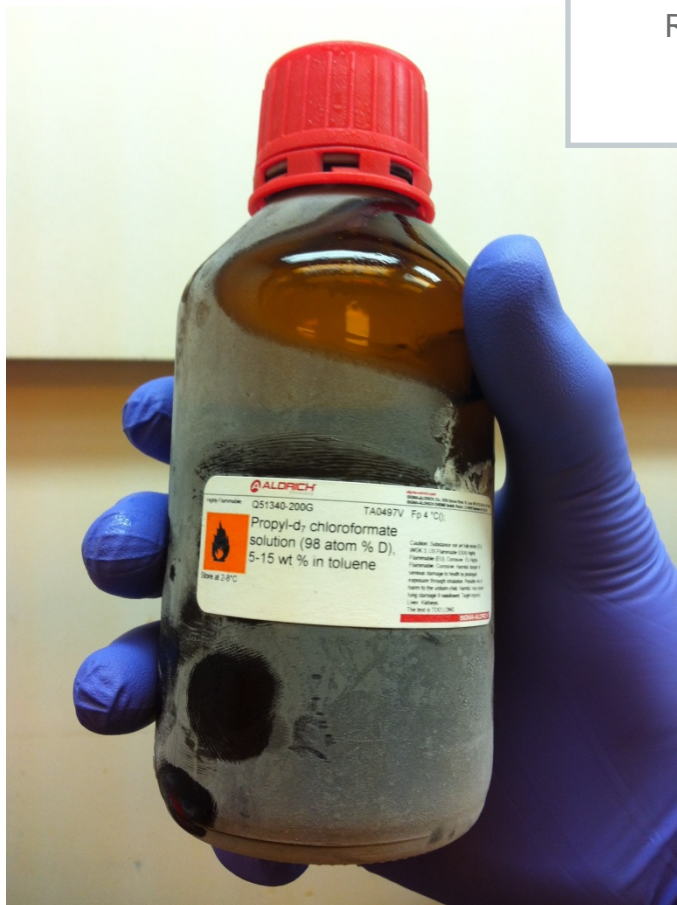
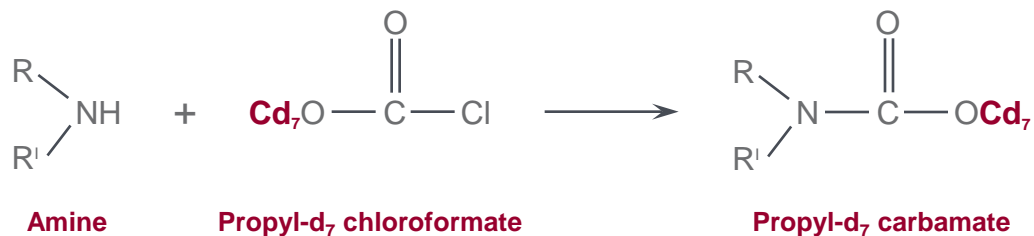
*Great assay*

## BAD NEWS

*No internal standard*

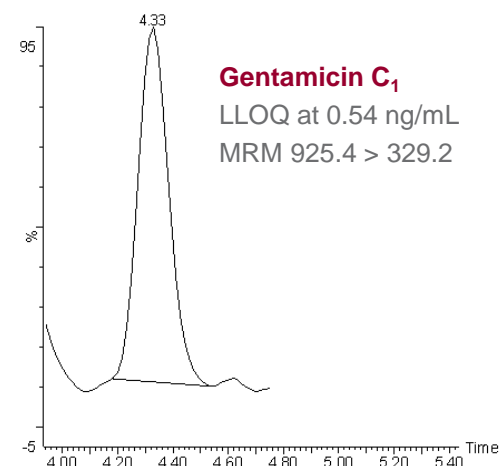
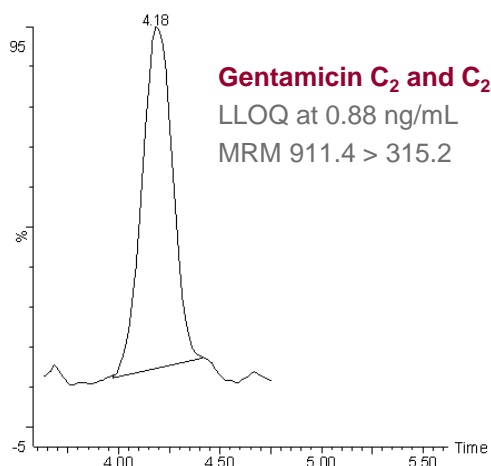
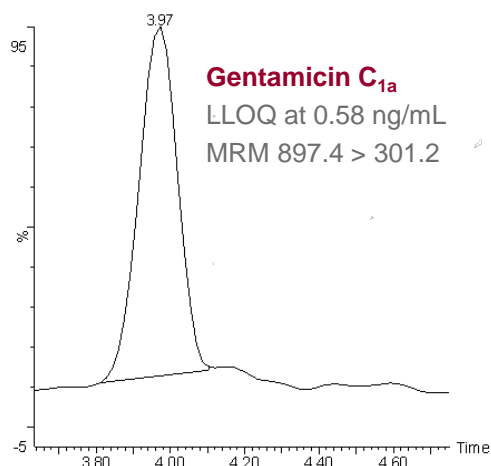
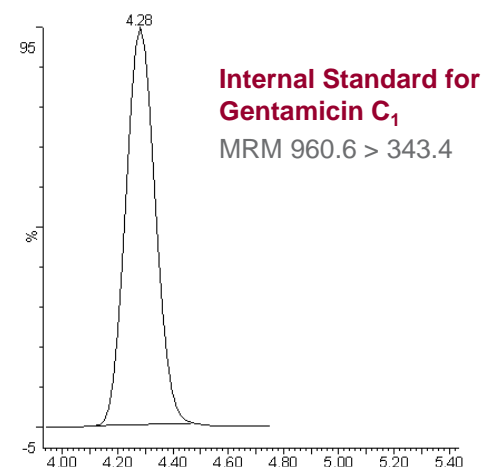
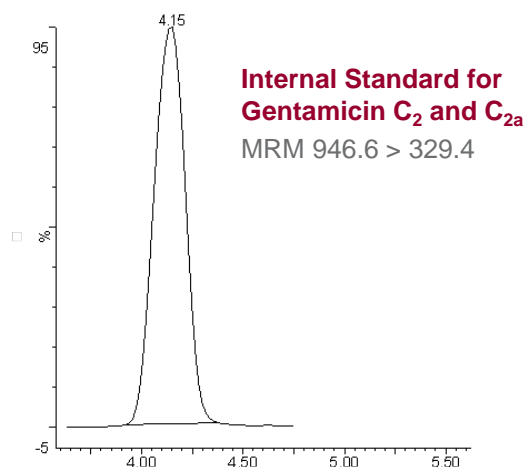
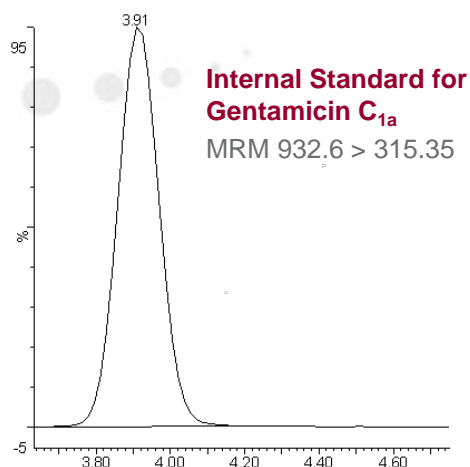


# The Solution



- Stable labeled I.S. is synthesized by pre-derivatization with d<sub>7</sub>-PCF
- I.S. is added at the beginning of extraction but is not derivatized *within* the assay
- Gentamicin isoforms in samples are derivatized within the assay by non-labeled PCF

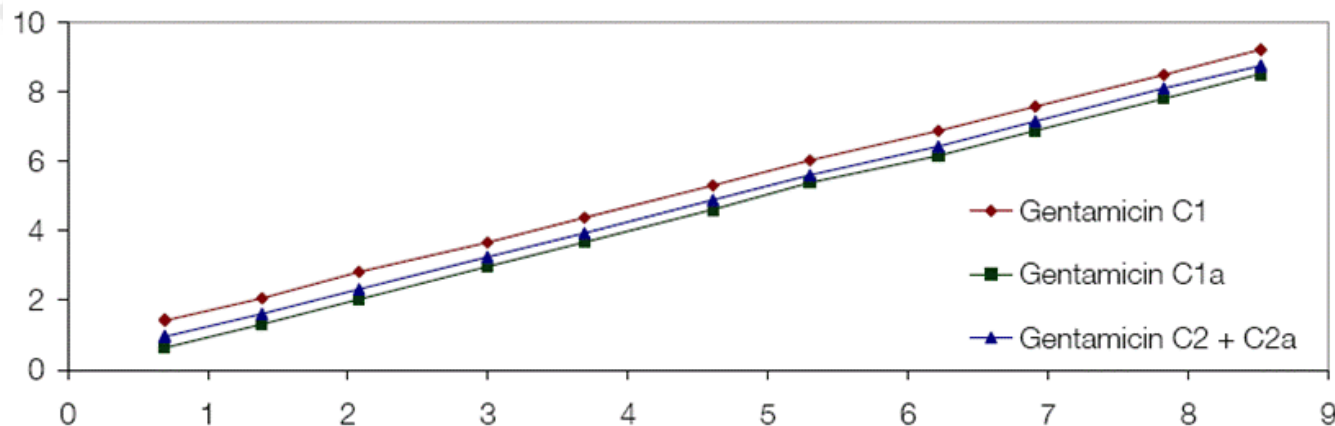
# Final Outcome



Note: Each isoform has its own I.S in the same relative abundance as the drug substance because they originate from the same reference material.

# Validation Summary

## Example Calibration Curves



## Interday Quality Control Sample Data and Statistics

	Gentamicin C <sub>1</sub> Concentration (ng/mL)			Gentamicin C <sub>1a</sub> Concentration (ng/mL)			Gentamicin C <sub>2</sub> + C <sub>2a</sub> Concentration (ng/mL)				
	1.62	54.0	1,080	1.74	58.4	1,180	2.64	88.0	1,760		
Mean	1.60	54.1	1,100	Mean	1.76	58.4	1,180	Mean	2.65	88.9	1,780
%CV	5.41	2.89	4.02	%CV	6.89	2.93	3.78	%CV	5.25	4.11	3.09
%DEV	-1.23	0.185	1.85	%DEV	1.15	0.690	1.72	%DEV	0.379	1.02	1.14

*Final validated range was 2.00 – 5,000 ng/mL total Gentamicin*

# Summary & Conclusions

- Novel application of deuterium labeled derivatization reagent to make an ideal I.S.
- Multiple benefits to chloroformate derivatization
- Critical aspects for success:
  - Parallel extraction for analyte and pre-derivatized I.S.
  - Robust derivatization – analyte has to “catch up” with I.S.
  - Analyte conversion – labeled derivative cannot exchange for unlabeled derivative
- Further applications:
  - General approach for other aminoglycoside antibiotics
  - Fluoro- $\beta$ -alanine (FBAL)
  - Doxorubicin and metabolite Doxorubicinol

# Acknowledgements

## ***Scientific support***

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